

NIPD GENETICS ForeSENT



PATIENT INFORMATION				REFERRAL INFORMATION			
NAME Jannis D	loe	CLINIC NAME Oncology Cente	CLINIC NAME Oncology Center				
Gender Male			CLINIC ID Clinic XXX				
DATE OF BIRTH 01/01/1965			REFERRING CL John Smith, ME	REFERRING CLINICIAN John Smith, MD			
TEST INDICATIONS Lung Adenocarcinoma			CLINIC EMAIL Clinicxxx@clinic	CLINIC EMAIL Clinicxxx@clinic.com			
SAMPL	E INFORMATION						
ORDER I NIPD12	NUMBER LAB NUMBER 34 1234TS		DATE OF COLLE 03/02/020	DATE OF COLLECTION DATE RECE 03/02/020 04/02/203			
CANCE	R TEST SELECTED						
	NTIA Pan-Cancer	able at the end of	the report				
TEST R	ESULTS						
POSITI At least	VE 1 clinically significa	int variant ide	ntified				
RESUL	T SUMMARY						
BIOMA	RKER FINDINGS						
MSI-H □ Dete	CTED 🛛 NOT-DE	ECTED					
GENON	AIC FINDINGS						
Gene	Variant Detected	Allele Fraction	FDA/EMA Approved Therapies (In patient's indications) *	FDA/EMA Approved Therapies (In other indications) **		Clinical Trials	
EGFR	c.2239_2256del (p.Leu747_Ser75 2del)	25.3%	 Afatinib and Dacomitinib and Erlotinib and Gefitinib and Osimertinib 	None		401	
TP53	c.893G>A (p.Arg298His)	12%	None	None		8	
Approv *List of FDA **List of FD/ indicated at Note: Clinica indicated fo	c.893G>A (p.Arg298His) red in indication and/or EMA approved drugs in A and/or EMA approved drugs i bove. al trials listed in this report are ir the indicated cancer type and	Lack of response the patient's cancer n other tumor types. T retrieved from clinical gene. The list of ther	Gefitinib Simertinib None Included in NCCN guidelin	es N guidelines for t Illing by invitation s report may not b	and recruiting trials be complete and/or	type are clo for the exhaustive	







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NIPD GENETICS ForeSENTIA tumor profile

instead it should be regarded as a supplementary source of information for guiding therapy decisions. All treatment decisions remain the full and final responsibility of the treating clinician.

INTERPRETATION

Variant summary:

EGFR c.2239_2256del (p.Leu747_Ser752del)

The EGFR variant c.2239_2256del is classified as Tier 1 variant with strong clinical significance. This variant is an in-frame deletion on exon 19 of the EGFR gene (NM_005228.4). This deletion results in a deletion of 6 amino acids at position 747-752 of the protein sequence. It has previously been identified in tissues derived from lung and submitted in the COSMIC database (COSMIC ID: COSM6255)⁵. EGFR encodes for the epidermal growth factor receptor with an important role in cell growth, proliferation and survival⁶. Somatic mutations in the tyrosine kinase domain of the (EGFR) gene are present in approximately 80% of the lung adenocarcinomas that respond to first and second generation EGFR inhibitors (eg, gefitinib, erlotinib and afatinib)⁷.

Several approved targeted therapies are associated with this alteration. An indicative list of available therapies approved for lung adenocarcinoma or in other cancer types is available in the results summary section. Please refer to the appropriate regulatory authority and guidelines for the selection of the appropriate therapy. Several clinical trials are active or currently recruiting patients diagnosed with non-small cell lung cancer with an EGFR mutation. Please refer to Clinicaltrials.gov for more information.

Guidelines from the National Comprehensive Cancer Network are available for patients diagnosed with nonsmall cell lung cancer with an EGFR mutation. Please refer to the NCCN guidelines for Non-small cell lung cancer for more information⁸.

TP53 c.893G>A (p.Arg298His)

The TP53 variant c.893G>A is classified as Tier 1 variant with strong clinical significance. This variant is a missence G>A substitution on exon 10 pf the TP53 gene (NM_001276760.1). This missense variant results in an arginine to Histidine amino acid substitution at position 298 of the protein sequence. This variant has been identified in tissues derived from liver, adrenal gland, skin, pancreas and cervix and submitted in the COSMIC database (COSMIC ID: COSM43882). TP53 encodes p53, a tumor suppressor protein that consists of transactivation domain, proline-rich domain, DNA-binding domain, oligomerization domain, and regulatory domain. p53 responds to diverse cellular stresses to maintain genomic stability and to induce cell cycle arrest, apoptosis, DNA repair and metabolic changes. TP53 mutations represent an important mechanism of resistance to DNA-damaging chemotherapeutic agents. Somatic TP53 mutations are found in a variety of cancers with various frequencies depending on cancer type; overall, TP53 is mutated in over one-half of human cancers. Missense mutations were the most frequent (~70-80%), followed by frameshift and nonsense mutations.

At the moment there are no approved therapies targeting tumors with TP53 alterations. TP53 mutations may be potential prognostic and predictive markers in some tumor types, as well as targets for pharmacological intervention in some clinical settings⁷. Please refer to Clinicaltrials.gov for more information.

VARIANTS OF UNKNOWN SIGNIFICANCE

MET(NM_001127500.3):c.3029C>T (p.Thr1010lle)

Note: One or more variants of unknown significance (VUS) have been detected in this patient's tumor sample. These variants are known as VUS due to their limited characterization and clinical evidence in the scientific literature at the time of writing of this report, making their significance unclear. However, we do include them here for reference in case they become clinically important in the future.







METHODOLOGY

ForeSENTIA is a Laboratory Developed Test (LTD) from NIPD Genetics Public Company Ltd for tumor molecular testing. Genomic deoxyribonucleic acid (gDNA) is extracted using a standardized methodology and subjected to enzymatic fragmentation and DNA library preparation. DNA enrichment for the genomic regions of interest is carried out using a solution-based hybridization method followed by next generation sequencing (NGS). Sequence data is aligned to a reference genome and variants are identified using proprietary bioinformatics pipelines. ForeSENTIA can be used for the identification of selected single nucleotide variants (SNVs), small insertions and deletions (Indels, \leq 30bp), translocations and copy number variations (CNAs) depending on the test ordered. Tumor-related actionable and clinically relevant alterations are reported. Analysis and Interpretation is performed using but not limited to Varsome Clinical CE-IVD platform (ISO 13485) according to published knowledge at the time of testing. Genetic counselling for the clinical interpretation and significance of the results is recommended. The ForeSENTIA test development and performance evaluation was carried out by NIPD Genetics Public Company Limited, which is regulated under the Clinical Laboratory Improvement Act of 1998 (CLIA) as qualified to perform high-complexity testing. ForeSENTIA is intended for clinical purposes and should not be regarded as investigational or for research. The test has not been cleared or approved by the U.S.Food and Drug Administration (FDA), which does not require this test to go through premarket FDA review.

TECHNICAL SPECIFICATIONS AND LIMITATIONS

This test aims to detect selected targeted variants, CNAs and translocations relevant to cancer development in the genes described above, by targeting the exons and hotspot regions in the genes described above. Variants that fall outside of the targeted bases are not intended to be detected by this assay. Sequence specific alterations such as SNVs and Indels are evaluated when the variant allele frequency exceeds 5% and 10% respectively. Translocations are reported when the frequency exceeds 20% and CNAs are evaluated when the copy number amplification is either \geq 4-fold with tumor DNA purity being >30% or \geq 6-fold with tumor DNA purity at 20%. Biallelic deletions in the genes tested for CNAs are evaluated when the tumor fraction is higher than 50%. The overall sensitivity and specificity of the assay is \geq 97% and 99.9%, respectively. Reduced sensitivity for the detection of the targeted genetic alterations may be due to low quality of the sample because of the procedure of tissue formalin-fixation or other factors that include, but are not limited to, low DNA yield, insufficient tumor DNA content and high intratumor heterogeneity in the specimen provided.

The test does not determine whether a variant is somatic or germline. Patients with an alteration identified in genes that are also associated with cancer predisposition might benefit from additional germline testing.

ADDITIONAL INFORMATION / DISCLOSURE

Test performance is valid only for the presence or absence of the tested cancer-associated variants in the genes included in the test. Therefore, a negative result indicates the absence of a cancer variant out of all the targeted variants included in the test and does not eliminate the possibility of a variant in a genomic position not tested by this assay. A positive result indicates the presence of a clinically relevant alteration. The results are interpreted based on information provided on the sample information form. Misinterpretation of results may occur if insufficient or inaccurate information is provided. A positive finding does not guarantee association with a certain treatment or drug. Drugs or treatments mentioned in this report may not necessarily be suitable for the patient. Decisions on medical management must be based on the clinician's judgment taking into consideration all available information such as the patient's medical history, family history and other medical tests and examinations performed.

Although this test is highly accurate, there is still a small possibility for false positive or false negative results. This may be caused by technical and/or biological limitations, including but not limited to poor sample quality, bone marrow transplants or other rare molecular events. Other reasons for false positive or false negative results include, but are not limited to: mislabeled samples, inaccurate reporting of clinical/medical information and rare technical errors.

Genetic testing is an important part of the diagnostic process. However, genetic tests may not always give a definitive answer. In some cases, testing may not identify a genetic variant even though one exists. This may be due to limitations in current medical knowledge or testing technology. Clinical correlation with other clinical data and tests is recommended. Results should always be considered in the context of other clinical criteria. The analysis is specific only for the test ordered. The referral clinician is responsible for counselling before and after







the test; including the provision of advice regarding the need for additional genetic testing. Other diagnostic tests may still be necessary.

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Elena Kypri, Ph.D. ASCP

Approved by:

Approved by:

Philippos Patsalis, Ph.D, HCLD, Laboratory Director

Date of report (DD/MM/YYYY): 10/07/2020









NIPD GENETICS ForeSENTIA tumor profile

	Fore	SENTIA PAN	CANCER Gen	e List	
AKT1 Exon 4 NM_001014432.1)	ALK Exons 18-20, 22-23 (NM_004304.5)	APC Full exonic coverage (NM_000038.6)	AR Full exonic coverage (NM_000044.6)	ARAF Exon 7 (NM_001654.5)	ATM Full exonic coverage (NM_000051.3)
			Exon 2 (NM_001011645.3)		
	Selected non-coding regions covered	Selected non-coding regions covered			Selected non-coding regions cover
					-
ATRX Exons 5, 7, 9, 11, 13-14, 17, 20-22, 29-31, 35 NM_000489.5)	BARD1 Full exonic coverage (NM_00465.4)	BRAF Exons 9-12, 15 (NM_001354609.2)	BRCA1 Full exonic coverage (NM_007294.4)	BRCA2 Full exonic coverage (NM_000059.3)	BRIP1 Full exonic coverage (NM_032043.3)
		Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions cover
					-
CDH1 Full exonic coverage NM_004360.5)	CDKN2A Full exonic coverage (NM_058195.3)	CHEK2 Exons 1-4, 6-11 (NM_001257387.2)	CIC Exon 19 (NM_015125.4)	CTNNB1 Exon 3 (NM_001904.4)	DDR2 Exon 17 (NM_006182.4)
	Selected non-coding regions covered	Selected non-coding regions covered			
DICER1 Exons 2-26	EGFR Exons 2-5, 8-10, 12, 14-15,	ERBB2 Exons 7-8, 11-12, 14, 23-28, 31	ERBB3 Exons 3, 6-11, 20, 21	ERBB4 Exons 7, 15, 19, 23	ESR1 Exons 1, 3-7, 9-10
NM_177438.2) Exons 1-3 NM_030621.4)	17-24, 27-28 (NM_005228.5)	(NM_001289936.1)	(NM_001982.3)	(NM_005235.3)	(NM_001122742.1) Exon 1 (NM_001291230.1)
Exon 1 (NM_001271282.3)					
Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions covered			Selected non-coding regions cover
			-	-	
EXW7 Exons 6-7, 9-14 NM_001349798.2)	FGFR1 Exons 3, 6-10, 13-14, 17, 19 (NM_001174064.2)	FGFR2 Exons 2-9, 11, 13-18 (NM_000141.4)	FGFR3 Exons 4, 7-8, 11-12, 16-18 (NM_000142.4)	FLT3 Exon 20 (NM_004119.3)	FOXA1 Exon 2 (NM_004496.5)
	Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions covered	A	A
OXL2	FUBP1	GATA3	GNA11	GNAQ	GNAS
xon 1 NM_023067.4)	Exon 14 (NM_003902.5)	Exons 5-6 (NM_001002295.2)	Exons 4-5 (NM_002067.5)	Exons 5, 7 (NM_002072.5)	Exon 8 (NM_000516.6)
	Selected non-coding regions covered				A
H3F3A	IDH1	IDH2	JAK2	KEAP1	KIT
xon 2 NM_002107.6)	Exon 4 (NM_005896.3)	Exon 4 (NM_002168.3)	Exon 14 (NM_004972.3)	Exons 2-4 (NM_203500.2)	Exons 2-21 (NM_000222.2)
					Selected non-coding regions cover
(RAS	MAP2K1	MAP3K1	MET	MLH1	MRE11A
xxons 2-4 NM_004985.5) xxon 5 NM_001369786.1)	Exons 2-3, 6-7 (NM_002755.3)	Exons 4-6, 16-18 (NM_5921.2)	Exons 2-6, 8-21 (NM_000245.4)	Full exonic coverage (NM_000249.3)	Exons 1-18, 20 (NM_005591.4)
elected non-coding regions covered			Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions cover
	A			A	
MSH2 Full exonic coverage NM_000251.3)	MSH6 Full exonic coverage (NM_000179.2)	MTOR Exons 43, 47, 53, 56 (NM_004958.4)	MYC Exons 1, 3 (NM_002467.6)	MYCN Exon 3 (NM_005378.6)	NBN Full exonic coverage (NM_002485.5)
Selected non-coding regions covered	Selected non-coding regions covered		Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions cover
			•	•	
NF1 :xons 6, 12, 17-18, 21-22, 25, 27-28, 34, 37, 40, 44-47, 19, 53 NM_001042492.3)	NPM1 Exon 11 (NM_002520.6)	NRAS Exons 2-4 (NM_002524.5)	NTRK1 Exons 7-15 (NM_002529.3)	NTRK2 Exons 12-13, 15-16 (NM_006180.4)	NTRK3 Exons 14-15 (NM_002530.4)
			Selected non-coding regions covered	Selected non-coding regions covered	Selected non-coding regions cove

Selected non-coding regions which are covered by the test are indicated above.

Single Nucleotide Variant / Indels

Copy Number Alterations

31 Neas Engomis Str, 2409 Engomi, Nicosia, Cyprus.Tel: + 357 22266888, Web Page: www.nipd.comPage: Web Page: Web Page: Web Page: Web Page: Web Page: Page

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Rearrangements





NIPD GENETICS ForeSENTIA tumor profile

PALB2 Full exonic coverage (NM_024675.4) Selected non-coding regions covered	PDGFRA Exon 18 (NM_006206.6)	PIK3CA Exons 2-6, 8, 10, 15-17, 19-21 (NM_006218.4) Selected non-coding regions covered	PIK3CB Exons 13, 15, 24 (NM_006219.3)	PMS2 Exons 6-8, 10 (NM_000535.7) Selected non-coding regions covered	POLE Exons 2-49 (NM_006231.4) Selected non-coding regions covered
A	A		A	A	A
PTEN Full exonic coverage (NH_000314.8) Selected non-coding regions covered	RAD51C Full exonic coverage (NH_058216.3) Selected non-coding regions covered	RAD51D Full exonic coverage (NH_002878.3) Selected non-coding regions covered	RAF1 Exon 7 (NM_001354689.3)	RB1 Exons 1-13, 16-27 (NM_0003212) Selected non-coding regions covered	RET Full exonic coverage (NM_020975.6) Selected non-coding regions covered
ROS1 Exons 31-36 (NM_002944.2) Selected non-coding regions covered	RUNX1 Exon 5 (NM_001754.4)	SMAD4 Full exonic coverage (NM_005359.6) Selected non-coding regions covered	SPOP Exons 4-5 (NM_001007228.2)	STK11 Full exonic coverage (NM_000455.5) Selected non-coding regions covered	TERT Exon 1 (NM_198253.3)
	A	A			A
TMPRSS2 Exons 1-4 (NM_005656.4) Selected non-coding regions covered	TP53 Full exonic coverage (NH_000546.6) Selected non-coding regions covered				

The pan-cancer panel also screens for 1p/19q codeletion

Selected non-coding regions which are covered by the test are indicated above.

Single Nucleotide Variant / Indels

Copy Number Alterations

Rearrangements







